

AbMax BiotechnoLogy Co., LTD
Address: 99Kechuang 14th Street,BuiLding 18,
Unit 2,Suite 201B,BDA,Beijing,
China Zip:101111
TeL: 86-10-59755729, Fax: 86-10-59755718
E-maiL:info@antibodychina.com
Website: www.antibodychina.com

HPV Type 31 In Vitro Potency Evaluation Kit Manual

Cat#17-0118

This kit is used to detect the content of HPV31-L1 antigen in samples.

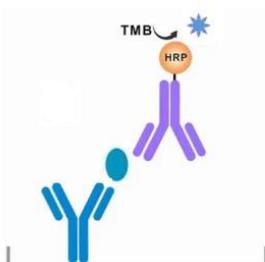
research use only and not for diagnostic purposes.

Table of Contents

1. Experimental Principle	错误！未定义书签。
2. Instructions for Use	错误！未定义书签。
3. Materials and Reagents Provided	错误！未定义书签。
4. Materials Required for the Experiment but Not Provided	1
5. Storage Conditions and Shelf Life	2
6. Operational Tips	2
7. Reagent Preparation	错误！未定义书签。
8. Reference and Sample Preparation	错误！未定义书签。
9. Flowchart	3
10. Operating Procedures	3
11. Result Processing	错误！未定义书签。
12. Product Performance Indicators	4
13. Problems and Solutions	7

1. Experimental Principle

This product uses the principle of a sandwich ELISA with a monoclonal neutralizing antibody against HPV31-L1 coated on the ELISA plate. After the addition of the sample and reaction, unbound materials are washed away. Another HRP-labeled monoclonal neutralizing antibody against HPV31-L1 is added, forming a complex of coated antibody-antigen-HRP antibody. The degree of TMB color development indicates the content of active HPV31-L1 antigen in the intermediate products of vaccine production, drug substance, and 9-valent HPV vaccine mixture (before adjuvant addition).



2. Instructions for Use

Please read this manual carefully before starting the experiment.

- Follow good laboratory practices: Wear gloves, lab coats, and safety goggles at all times. Do not eat, drink, or smoke in the laboratory area.
- All biological materials should be treated as potentially hazardous and disposed of according to established safety procedures.
- Do not mix or substitute reagents or materials from other kit batches or suppliers; performance cannot be guaranteed if substituted.

3. Materials and Reagents Provided

Components of the kit	Quantity	Storage Conditions
Pre-coated plate	8 wells × 12 strips	2~8°C
Detection antibody (500×)	50μL × 1 tube	-15 ~ -25°C
Single component color development solution II	11mL × 1 bottle	2~8°C
Termination solution	7mL × 1 bottle	2~8°C
BSA	3g/bag × 2 bags	2~8°C
Detection antibody diluent	12mL × 1 bottle	2~8°C
20×PBST	50mL × 1 bottle	2~8°C
Sealing membrane	2 sheets	2~8°C/Room temperature
Manual	1 copy	2~8°C/Room temperature

4. Materials Required for the Experiment but Not Provide

These materials are not included in the kit but are needed for the test:

- 1) HPV31 antigen reference solution
- 2) HPV31 antigen enterprise reference solution

3) 9-valent HPV mixture enterprise reference solution (optional)

- Microplate reader, deionized water, multi-channel and single-channel pipettes, dilution tubes, shaker incubator.

5. Storage Conditions and Shelf Life

- Detection antibody (500×), store at -15 ~ -25°C, other components at 2~8°C away from light.
- Validity period of 12 months.

6. Operational Tips

- Avoid creating bubbles when preparing solutions or adding samples.
- Replace tips promptly to avoid cross-contamination of samples or reagents.
- Ensure the ELISA plate is covered with a sealing membrane during incubation.
- All samples should be thoroughly and gently mixed.
- Incubate the ELISA plate on a shaker incubator during all incubation steps.

7. Reagent Preparation

- Equilibration: Bring the required reagents to room temperature (18 ~ 25°C) and equilibrate for 30 minutes.
- 20×PBST may contain precipitates, which is normal. If the precipitate does not dissolve by gentle stirring, it can be heated in a 37°C oven to dissolve the precipitate before use.

7.1 Preparation of 1×PBST

Take 1 bottle of 20×PBST and dilute with deionized water to 1000mL, mix well and set aside for use. The amount of solution can be adjusted according to the experimental needs, prepare as needed.

7.2 Preparation of 5% BSA diluent

Accurately weigh 5.0g of BSA and completely dissolve it in 100mL of 1×PBST, mix well and set aside for use as 5% BSA diluent.

The amount of solution can be adjusted according to the experimental needs, prepare as needed.

7.3 Preparation of enzyme conjugate-detection antibody working solution

Calculate the volume of working solution needed for the experiment, take an appropriate amount of enzyme conjugate (500×) and dilute with detection antibody diluent 100 times, then further dilute 5 times, mix well and set aside for use.

8. Reference and Sample Preparation

- Prepare different concentrations of reference solutions (2-fold gradient dilution) with 5% BSA diluent: 4000ng/mL, 2000ng/mL, 1000ng/mL, 500ng/mL, 250ng/mL, 125ng/mL, 62.5ng/mL, 31.25ng/mL, 15.625ng/mL, 7.8125ng/mL, 3.9ng/mL, 0ng/mL (5% BSA diluent).
- Dilute the test samples with 5% BSA diluent to the linear range of the standard curve for detection. If the concentration of the test sample is unknown, it can be diluted 10 times, 30 times, 100 times, 300 times, and 1000 times, and then tested to select the OD value within the standard curve range to back-calculate the sample concentration (ng/mL). Finally, the sample

concentration should be multiplied by the sample dilution factor to obtain the measured value.

- Prepare new reference and sample solutions each time, and discard used reference and sample solutions.

9. Flowchart

9.1 Prepare all reagents, samples, and reference standards according to the manual.

9.2 Add reference standards or samples (100 μ L) into the wells of the microplate, incubate at 37°C with shaking for 60 minutes.

9.3 Wash with 1x PBST three times.

9.4 Add the enzyme-labeled reagent (100 μ L), and incubate at 37°C for 60 minutes.

9.5 Wash with 1x PBST three times.

9.6 Add the color development solution (100 μ L), and let it react at room temperature in the dark for 10 minutes.

9.7 Add 50 μ L of the stopping solution to all wells, and read the OD values.

10. Operating Procedures

- Prepare all reagents, references, and samples according to the requirements of the previous sections.

- The ELISA plate is ready to use at any time, and it is not necessary to rinse before use.

10.1 Sample Addition

Add all references and samples to the ELISA plate, mark them, add 100 μ L of sample per well (set up duplicate wells), seal the plate with a sealing membrane, and place it in a 37°C constant temperature shaker incubator, 300rpm, incubate for 60 minutes.

10.2 Washing

Discard the liquid in each well, add 350 μ L of 1 \times PBST to each well, let it stand for 30 seconds, discard the liquid, and repeat 3 times. After the last wash, gently blot dry on a paper towel.

10.3 Addition of Enzyme Conjugate

Add 100 μ L of enzyme conjugate working solution to each well, seal the plate with a new sealing membrane, and place it in a 37°C constant temperature shaker incubator, 300rpm, incubate for 60 minutes.

10.4 Washing

Repeat step 10.2.

10.5 Color Development

Add 100 μ L of single component color development solution to each well, gently mix and incubate in the dark at 25°C for 10 minutes.

10.6 Termination

Add 50 μ L of termination solution to each well, gently mix.

10.7 Reading

Select the main wavelength of the microplate reader at 450nm, reference wavelength at 630nm, and measure the absorbance value (OD value=OD₄₅₀-OD₆₃₀) of each well.

11. Result Processing

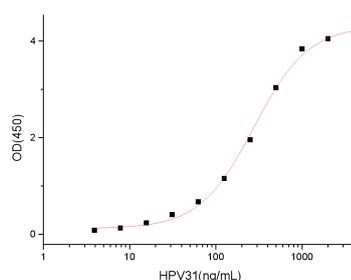
11.1 Standard Curve

Calculate the average OD value of the reference (see example below, for illustration only,

specific to actual measurement):

Standard Curve ng/mL	OD1	OD2	Average OD
4000	4.024	4.243	4.134
2000	4.068	4.032	4.050
1000	3.735	3.939	3.837
500	2.934	3.145	3.040
250	1.933	1.986	1.960
125	1.174	1.143	1.159
62.5	0.700	0.654	0.677
31.25	0.414	0.399	0.407
15.625	0.231	0.238	0.235
7.8125	0.13	0.13	0.130
3.90625	0.08	0.085	0.083
NC	0.036	0.036	0.036

Fit the standard curve concentration and corresponding average OD values using a four-parameter fitting method to obtain the standard curve, as shown in the figure below. The four-parameter fitting equation is: $y = 4.31597 - 4.19815 / [1 + (x/276.77958)^{1.37593}]$.



Four-parameter fitting standard curve (linear correlation coefficient $R^2=0.99726$)

11.2 Calculation of Sample Concentration

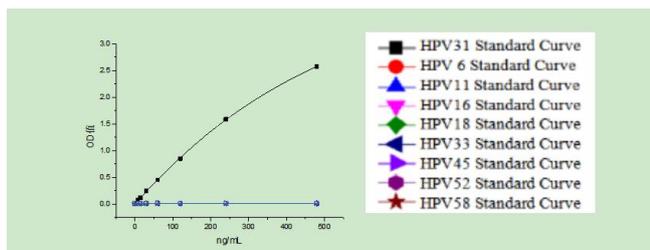
Select the OD value within the standard curve range to back-calculate the sample concentration, which is the measured value (ng/mL). Multiply the measured value by the sample dilution factor to obtain the sample concentration.

12. Product Performance Indicators

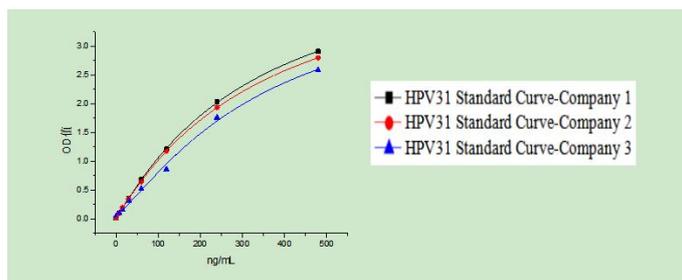
- Linear range: 3.9 ~ 4000ng/mL
- Sensitivity: <3.9ng/mL.
- Accuracy and precision: The recovery rate of high, medium, and low concentration quality control points is within the range of 80% ~ 120%, which meets the requirements; CV (%) ≤ 10%, which meets the requirements.

Accuracy/Precision	Accuracy (Recovery Rate %)			Precision (CV %)		
	1	2	3	1	2	3
Experiment						
HQC-400ng/mL	95%	102%	106%	1.82%	1.56%	3.27%
MQC-100ng/mL	95%	94%	103%	2.01%	1.42%	4.07%
LQC-20ng/mL	100%	96%	100%	1.36%	2.02%	2.90%

- HOOK effect: Using HPV31 reference solution higher than 4000ng/mL for testing, the signal value did not decrease, no HOOK effect.
- Dilution linearity: Select reference solutions higher than 4000ng/mL diluted to the linear range of the standard curve (400ng/mL, 100ng/mL, 20ng/mL), calculate accuracy, and the deviation is within the range of 80% ~ 120%.
- Specificity: No significant cross-reactivity with other 8 HPV subtypes such as HPV6, 11, 18, 16, 33, 45, 52, 58.



- Broad-spectrum Activity: It shows binding affinity with HPV31 from more than three different companies with varying expression systems.

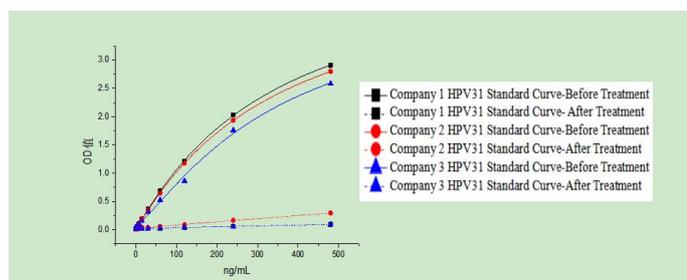


- Applicability: Suitable for detecting the content of active HPV31 antigen in both HPV31 monovalent solutions and 9-valent HPV mixtures (before adjuvant addition). There is no cross-reactivity with other eight HPV subtypes in vaccines other than HPV31, with a total cross-reactivity rate of 0%. The OD values detected for HPV31 in the 9-valent mixture are close to those detected for the monovalent, with the standard curves essentially overlapping, making it suitable for the detection of HPV31 in 9-valent HPV vaccines.

Sample	HPV31				8 mix-HPV 6、18、33、45、52、16、11、58				9 mix-HPV6、11、31、33、45、52、16、18、58			
	OD	OD	Ave OD	P/N	OD	OD	Ave OD	P/N	OD	OD	Ave OD	P/N
480	2.324	2.337	2.331	40.89	0.061	0.059	0.060	1.09	2.255	2.233	2.244	44.00
240	1.410	1.353	1.382	24.24	0.050	0.060	0.055	1.00	1.269	1.324	1.297	25.42
120	0.821	0.740	0.781	13.69	0.044	0.050	0.047	0.85	0.682	0.723	0.703	13.77
60	0.445	0.429	0.437	7.67	0.048	0.049	0.049	0.88	0.411	0.384	0.398	7.79
30	0.236	0.242	0.239	4.19	0.043	0.046	0.045	0.81	0.216	0.218	0.217	4.25
15	0.139	0.150	0.145	2.54	0.045	0.050	0.048	0.86	0.134	0.129	0.132	2.58

7.5	0.101	0.103	0.102	1.79	0.050	0.045	0.048	0.86	0.091	0.092	0.092	1.79
NC	0.057	0.074	0.057	1.00	0.059	0.051	0.055	1.00	0.051	0.051	0.051	1.00

- **Vaccine Inactivation Treatment:** After treatment at 56°C for 30 minutes, the HPV31 antigen solutions from three different companies showed a decrease in signal compared to untreated samples, indicating that this kit can reflect whether the HPV antigen solution has been inactivated.



- **Vaccine Testing:**
 - Recommended desorption method is as follows:
 - ◆ Preparation of Desorption Agent: 60mM sodium dihydrogen phosphate, 0.1M sodium citrate, 1M sodium chloride, 0.8% Tween-80 (pH 6.7-6.8).
 - ◆ Desorption Treatment of Finished Vaccine: Mix the desorption agent with the finished vaccine in a 1:1 ratio, then stir overnight at room temperature to react.
 - ◆ Desorption Efficiency (Measured Value/Theoretical Value):

QCs, ng/mL	Desorption Efficiency%
100	72%
50	75%
20	69%

- **Good Reproducibility:** After desorption of the finished vaccine, the coefficient of variation (CV) of the optical density (OD) values for each point on the standard curve is within 10%, and the CV of the OD values for quality controls (QCs) is also within 10%, indicating stable repeatability of the test.

Opration	Standard -9-valent mixed bulk solution						After desorption treatment of the finished vaccine							
	standard g/mL	OD1	OD2	Ave OD	QCs, ng/mL	OD	Ave OD	OD1	OD2	Ave OD	QCs, ng/mL	OD	Ave OD	ng/mL
500	4.006	4.01	4.008	100	3.016	2.893	3.808	3.747	3.778	100	2.79	2.727	71.642	72%
250	3.78	3.685	3.733		2.829		3.465	3.438	3.452		2.649			
125	3.248	3.255	3.252		2.835		3.048	2.94	2.994		2.743			
62.5	2.644	2.553	2.599	50	2.21	2.151	2.262	2.34	2.301	50	2.091	2.046	37.518	75%
31.25	1.85	1.798	1.824		2.096		1.573	1.589	1.581		2.025			
15.625	1.218	1.176	1.197		2.146		0.998	1.029	1.014		2.022			
7.8125	0.759	0.757	0.758	20	1.232	1.254	0.591	0.585	0.588	20	1.085	1.101	13.757	69%

NC	0.079	0.078	0.079		1.247		0.084	0.076	0.08		1.101			
					1.283						1.117			

13. Problems and Solutions

Problem	Cause	Solution
Poor standard curve linearity	Improper dilution of the standard curve	Check the pipette, use a calibrated pipette, standardize operations, and redilute.
Low signal value	Insufficient incubation time	Ensure adequate incubation time.
	Insufficient reagent amount or improper dilution	Check the pipette, ensure correct and sufficient reagent preparation
	Prolonged exposure of color development solution	The color development solution should be stored away from light, replace with a new color development solution.
High CV value	Inadequate washing	Follow the washing steps strictly.
	Contaminated washing solution	Replace with a new washing solution.
Insufficient sensitivity	Improper storage of the ELISA kit	Store according to the manual instructions
Precipitation in the diluent	Precipitation and/or coagulation of components in the diluent	Heat the diluent to 37°C to completely dissolve the precipitate.